

repliQa HiFi ToughMix®

Cat No. 95200-025 Size: 25 x 25 µL reactions (1 x 0.625 mL)

95200-100 100 x 25 μL reactions (1 x 2.50 mL) 95200-500 500 x 25 μL reactions (1 x 12.50 mL)

Description

The repliQa HiFi ToughMix is a unique, next generation 2x master mix that has 90x higher fidelity compared to Taq. The ToughMix has extreme speed, with extension times as fast as 1-10 kb/sec depending on target length. Additionally, the ToughMix has long range amplification properties as it can amplify hgDNA targets up to 24 kb and lambda targets up to 40kb.

Store at -25°C to - 15°C

The ToughMix is formulated with a genetically modified DNA polymerase coupled with hot start antibodies. It has $5' \rightarrow 3'$ polymerase activity, $3' \rightarrow 5'$ exonuclease activity, and generates blunt-ended products while providing the ability to amplify through uracils and primers containing inosines/uracils. It is tough tested, and is tolerant to multiple PCR inhibitors.

Components

repliQa HiFi ToughMix 2x reaction buffer containing optimized concentrations of MgCl₂,

dNTP's and proprietarily formulated HiFi polymerase,

hot start antibodies and ToughMix chemistry

Storage and Stability

Store kit components in a constant temperature freezer at -25°C to -15°C protected from light upon receipt. For lot specific expiry date, refer to package label, Certificate of Analysis or Product Specification Form.

Guidelines for PCR

- The design of highly specific primers is a critical parameter for successful PCR. The use of computer aided primer design programs is encouraged in order to minimize the potential for internal secondary structure. For best results, primer size should be limited to 22 35 bp with a melting point of at least 63°C. Ideal GC-content of the primers is 45-60%. A final concentration of 300 nM each primer is effective for most applications. Primers with inosine (dl) and uracil (dU) are acceptable.
- Preparation of a reaction cocktail is recommended to reduce pipetting errors and maximize assay precision. Assemble the reaction cocktail with all required components except sample template and dispense equal aliquots into each reaction tube. Add the DNA template to each reaction as the final step. Addition of samples as 2 to 5 μL volumes will improve assay precision.
- Suggested input quantities of template are: genomic DNA ≤ 200 ng; plasmid DNA ≤ 50 ng; cDNA ≤ 750 ng.
- After sealing each reaction, vortex gently to mix contents. Centrifuge briefly to collect components at the bottom of the reaction tube.
- Longer targets may require a higher primer melting temperature of at least 65°C, and a lower primer concentration
 of 150 nM.

Reaction Assembly

Component	Volume for 50 μL rxn	Final Concentration
repliQa HiFi ToughMix (2X)	25 μL	1x
Forward primer	variable	300 nM
Reverse primer	variable	300 nM
Nuclease-free water	variable	
Template	2 – 5 μL	variable
Final Volume (µL)	50 μL	

Note: For smaller or larger reaction volumes scale all components proportionally.

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PCR Cycling Protocol

PCR cycling (25 - 45 cycles):

*We recommend using 2-step cycling first. If you are have having difficulty, we suggest trying 3-step cycling.

2-Step Cycling*	3-Step Cycling	
98°C, 10s	98°C, 10s	
	(Tm -5) °C, 5s	
68 ° C	68 ° C	
≤ 1 kb: 1 sec	≤ 1 kb: 1 sec	
1 ~ 10 kb: 5 sec/ kb	1 ~ 10 kb: 5 sec/ kb	
≥ 10 kb: 10 sec/ kb	≥ 10 kb: 10 sec/ kb	

Quality Control

Kit components are free of contaminating DNase and RNase. 2x repliQa HiFi ToughMix is functionally tested for amplification of a 4-kb fragment from a single-copy gene in a human genomic DNA.

Nuclease Assay:

DNase: DNase activity must be below the detectable limits of 100 pg DNase I equivalent as assayed using a fluorogenic substrate following a 1 hour incubation at 37°C with each kit component at 1X concentration.

RNase: RNase activity must be below the detectable limits of 1 pg RNase A equivalent as assayed using a fluorogenic substrate following a 1 hour incubation at 37°C with each kit component at 1X concentration.

4.1 kb PCR Functional Assay: Negative control must be free of visible product with a single band at ~4.1 kb visible from 35 cycles of PCR using 20 ng human genomic DNA.

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