

## Moxi GO II - Mitochondrial Membrane Potential TMRE **Staining**

## Overview:

Tetramethylrhodamine, ethyl ester (TMRE) is a cell-permeant fluorescent dye that, due to it's cationic nature, is readily sequestered by active mitochondria. TMRE has a peak excitation of 549nm but can be excited by either the 488nm or 532nm laser. The peak emission is 574nm, requiring the use of the 561nm/LP filter on the Moxi GO II instruments.

## **Reagents/Components:**

- Orflo Moxi GO II Next Generation Flow Cytometer (Orflo Cat #MXG102)
- MF-S+ Cassettes (Orflo Cat #MXC030)
- Tetramethylrhodamine ethyl ester perchlorate (e.g. Sigma Cat #87917)
- Cell Staining Buffer (BioLegend cat # 420201 or PBS with 0.5% BSA, 0.1% Azide). *Note:* Stock PBS (any formulation, e.g. Gibco, Cat #10010023) can be used but might have slightly lower signal-to-noise ratios (higher background) for the assay.
- DMSO (any brand, e.g. Sigma #D8418)
- Optional/Recommended: Orflo Flow Reagent (Orflo Cat #MXA080)

## **Mitochondria - TMRE Labeling Protocol**

- 1. Make initial stock solutions of TMRE by sequential dilution of TMRE (Sigma Cat #87917) as follows:
  - a. 10mM stock TMRE: 25mg dissolved TMRE in 4.85ml DMSO
  - b. 10µM stock TMRE: 10µL of 10mM TMRE stock in 990µL DSMO
- 2. Dilute cells to a concentration of ~2e5-3e5 cells/ml with Cell Staining Buffer. *Notes:* 
  - a. Cells can be labeled directly in in culture media if they are already at the correct concentration (or are under-concentration).
  - b. Cells need to be in a single cell suspension for testing. Detachment with Accutase/Accumax is recommended with pipette trituration to break apart clusters
- 3. Aliquot 500μL of cell suspensions (it is useful to generate an FCCP-treated (i.e. 100μM), Sigma Cat #C2920, sample as a negative control) into separate polypropylene microcentrifuge tubes (Santa Cruz, Cat #sc-200271) – Note: do not use polystyrene (PS) as TMRE can bind significantly to PS.
- 4. Add 2.5μL of 10μM TMRE stock solution to each vial to achieve a 50nM final TMRE concentration and gently vortex to disperse. NOTE: At high overly high concentrations, TMRE has a quenching effect. Consequently, optimal TMRE concentration is dependent on cell type and sample prep and can vary from 20nM -200nM. Initial titration tests should be performed for optimal concentrations. As an initial guess, start with 50nM TMRE.
- 5. Incubate TMRE/Cell media at 37°C for 15-30 minutes in the dark.
- 6. Post-incubation, incubate cells for an additional 15min (RT/Dark).
- 7. Optional/Recommended: Add 20µL Orflo Flow Reagent per ml of cells and inversion mix sample.
- 8. Immediately run the samples on the Moxi GO II system using the "Open Flow Cytometry" assay with the "Medium" gain setting.

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